NOTE: Please save this file locally before filling in the table, DO NOT work on the file within your internet browser as changes will not be saved. Adobe Acrobat Reader (available free here) is recommended for completion.

The ARRIVE guidelines 2.0: author checklist

The ARRIVE Essential 10

These items are the basic minimum to include in a manuscript. Without this information, readers and reviewers cannot assess the reliability of the findings.

Item		Recommendation	Section/line number, or reason for not reporting
Study design	1	For each experiment, provide brief details of study design including: a. The groups being compared, including control groups. If no control group has been used, the rationale should be stated. b. The experimental unit (e.g. a single animal, litter, or cage of animals).	1a: Materials and Methods, Section 2.3 (Experimental Groups), Table 1. 1b: Materials and Methods, Sections 2.8 (Histopathological Examination) and 2.9 (Immunohistochemical Examination); the individual animal was the experimental unit.
Sample size	2	a. Specify the exact number of experimental units allocated to each group, and the total number in each experiment. Also indicate the total number of animals used. b. Explain how the sample size was decided. Provide details of any *a priori* sample size calculation, if done.	2a: Materials and Methods, Section 2.3 (Experimental Groups), and Table 1: Each experimental unit was an individual rat. Six rats were allocated to each of four groups, giving a total of 24 rats used in the experiment. Materials and Methods, Section 2.3 (Experimental Groups): Group size was specified in the ethics-approved protocol with consideration of the Reduction principle of the 3Rs. No formal a priori sample-size or power calculation was performed.
Inclusion and exclusion criteria	3	a. Describe any criteria used for including and excluding animals (or experimental units) during the experiment, and data points during the analysis. Specify if these criteria were established *a priori.* If no criteria were set, state this explicitly. b. For each experimental group, report any animals, experimental units or data points not included in the analysis and explain why. If there were no exclusions, state so. c. For each analysis, report the exact value of *n* in each experimental group.	3a: Materials and Methods, Section 2.1 (Animals and Housing): Only rats with a regular 4–5-day estrous cycle during acclimatization were included. No additional exclusion criteria are reported. 3b: Results, Section 3 (opening paragraph): No animal deaths or exclusions occurred; all 24 rats were included in the final analyses. 3c: Materials and Methods, Section 2.3 (Experimental Groups), and Tables 2–5: n = 6 animals per group.
Randomisation	4	a. State whether randomisation was used to allocate experimental units to control and treatment groups. If done, provide the method used to generate the randomisation sequence. b. Describe the strategy used to minimise potential confounders such as the order of treatments and measurements, or animal/cage location. If confounders were not controlled, state this explicitly.	4a: Materials and Methods, Section 2.3 (Experimental Groups): Animals were assigned to the four groups using a computer-generated random sequence. 4b: Materials and Methods, Sections 2.8 and 2.9: Housing conditions were standardized, and coded histology and immunohistochemistry slides were assessed in random order. The order of treatment and surgery and the distribution of groups across cages are not reported.
Blinding	5	Describe who was aware of the group allocation at the different stages of the experiment (during the allocation, the conduct of the experiment, the outcome assessment, and the data analysis).	Materials and Methods, Sections 2.8 and 2.9 An experienced histopathologist evaluated coded histopathology and immunohistochemistry slides in random order while blinded to group assignment. Blinding during group allocation, experimental procedures, biochemical measurements, and data analysis is not reported.
Outcome measures	6	a. Clearly define all outcome measures assessed (e.g. cell death, molecular markers, or behavioural changes). b. For hypothesis-testing studies, specify the primary outcome measure, i.e. the outcome measure that was used to determine the sample size.	6a: Materials and Methods, Sections 2.3 and 2 6b: Materials and Methods, Section 2.3: No single primary outcome was prespecified, and no a priori sample-size calculation was performed.
Statistical methods	7	a. Provide details of the statistical methods used for each analysis, including software used. b. Describe any methods used to assess whether the data met the assumptions of the statistical approach, and what was done if the assumptions were not met.	7a: Materials and Methods, Section 2.10 (Statistical Analysis): Statistical tests, pairwise comparisons, multiple-comparison adjustment, and software are reported separately for biochemical outcomes and histopathological/immunohistochemical 7b: Materials and Methods, Section 2.10: Normality was assessed with the Shapiro–Wilk test and variance equality with Levene's test. Animal-level histopathological and immunohistochemical scores were assessed with Kruskal–Wallis and exact
Experimental animals	8	a. Provide species-appropriate details of the animals used, including species, strain and substrain, sex, age or developmental stage, and, if relevant, weight. b. Provide further relevant information on the provenance of animals, health/immune status, genetic modification status, genotype, and any previous procedures.	8a: Materials and Methods, Section 2.1 (Animals and Housing): Twenty-four adult female Wistar albino rats aged 10–12 weeks and weighing 225–250 g were used. The substrain is not reported. 8b: Materials and Methods, Section 2.1: The animals were obtained from the Experimental Animals Application and Research Center of Erzincan Binali Yıldırım University. Health/immune status and previous procedures are not reported.
Experimental procedures	9	For each experimental group, including controls, describe the procedures in enough detail to allow others to replicate them, including: a. What was done, how it was done and what was used. b. When and how often. c. Where (including detail of any acclimatisation periods). d. Why (provide rationale for procedures).	9a: Materials and Methods, Sections 2.3–2.10 and Table 1: Drugs, doses, sham and CLP procedures, anesthesia, sample collection, and outcome measurements are described for each group. 9b: Materials and Methods, Sections 2.3.4 and 2.5: Injections were given once daily for seven days before surgery; samples were collected 24 hours after sham surgery or CLP. 9c: Materials and Methods, Sections 2.1 and 2.7.1. The seven-day acclimatisation period and housing conditions are described in Section 2.1. The locations of the laboratory analyses are reported in Section 2.7.1. 9d: Introduction, final paragraph, and Materials and Methods, Section 2.3 and 2.5: The study aim and the rationale for the CZA dose and seven-day exploratory pretreatment regimen are stated.
Results	10	For each experiment conducted, including independent replications, report: a. Summary/descriptive statistics for each experimental group, with a measure of variability where applicable (e.g. mean and SD, or median and range). b. If applicable, the effect size with a confidence interval.	10a: Results, Sections 3.1–3.3 and Tables 2–5: Results for each of the four groups are reported as mean ± standard deviation (n = 6 animals per group). 10b: Effect sizes with confidence intervals are not reported.